



Agilent Seahorse XFp Real-Time ATP Rate Assay Kit

User Guide **Kit 103591-100**

For use with Agilent Seahorse XFp Extracellular Flux Analyzers

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Assay Background

The cellular rate of adenosine triphosphate (ATP) production is a highly informative measurement to describe cellular metabolism, as ATP is the ubiquitous, dominant energy currency for cells. Cellular metabolic regulation allows cells to adjust for changes in ATP demand with subsequent changes in ATP production to maintain total intracellular ATP levels.

The Agilent Seahorse XFp Real-Time ATP Rate Assay is designed to measure total ATP production rates in living cells. Even more relevant is the ability of this assay to distinguish between the fractions of ATP that are produced from mitochondrial oxidative phosphorylation (OXPHOS) and glycolysis, the two main metabolic pathways responsible for ATP production in mammalian cells.

The Seahorse XFp Real-Time ATP Rate Assay uses metabolic modulators (oligomycin and a mix of rotenone and antimycin A, see **Figure 1** on page 6) that when serially injected, allow the calculation of the mitochondrial and glycolytic ATP production rates. Together with a Seahorse XFp Analyzer, the Seahorse XFp Real-Time ATP Rate Assay provides a new dynamic and quantitative insight into cellular bioenergetics by providing a real-time measurement of cellular ATP production rates and a quantitative phenotype of cellular energy poise.

1 Introduction

Assay Background

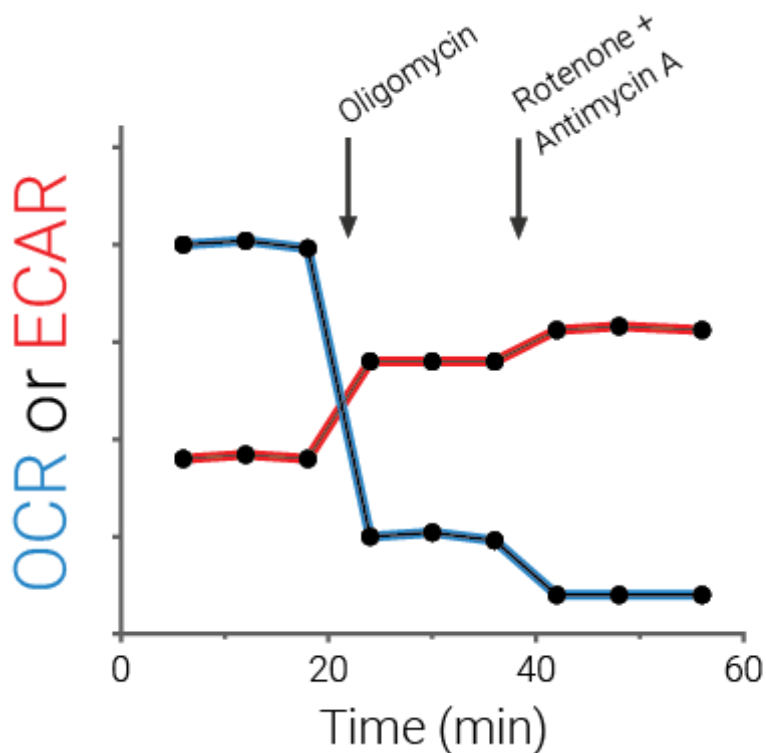


Figure 1. Representative scheme of Agilent Seahorse XFp Real-Time ATP Rate Assay - Kinetic profile of OCR and ECAR measurements. Basal OCR and ECAR rates are first measured. Injection of oligomycin results in an inhibition of mitochondrial ATP synthesis that results in a decrease in OCR, allowing the mitoATP Production Rate to be quantified. ECAR data combined with the buffer factor of the assay medium allows calculation of total Proton Efflux Rate (PER). Complete inhibition of mitochondrial respiration with rotenone plus antimycin A allows accounting for mitochondrial-associated acidification, and when combined with PER data allows calculation of the glycoATP Production Rate.

In mammalian cells, glycolysis and oxidative phosphorylation (OXPHOS) pathways provide the majority of cellular ATP. While OXPHOS consumes O_2 , driving the oxygen consumption rate (OCR), both pathways can contribute to the acidification of the assay medium. Conversion of glucose to lactate through glycolysis is accompanied by extrusion of one H^+ per lactate, while the TCA cycle that fuels ETC/OXPHOS produces CO_2 , which also results in acidification of the assay medium. The sum of these reactions is the primary driver of changes in extracellular acidification (ECAR).

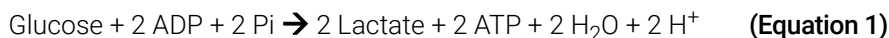
1 Introduction

Glycolytic ATP production rate calculation

Seahorse XF technology measures the flux of both H⁺ production (ECAR) and O₂ consumption (OCR), simultaneously. By obtaining these data under basal conditions and after serial addition of mitochondrial inhibitors (oligomycin and rotenone/antimycin A), total cellular ATP Production Rates and pathway-specific mitoATP and glycoATP Production Rates can be measured label-free and in real time. The series of calculations used to transform the OCR and ECAR data to ATP Production Rates is described below and are performed post data acquisition using the Seahorse XF Real-Time ATP Rate Assay Report Generator.

Glycolytic ATP production rate calculation

During conversion of one molecule of glucose to lactate in the glycolytic pathway, two molecules each of ATP, H⁺ and lactate are produced (**Equation 1**):



Considering the stoichiometry of the glycolytic pathway (**Equation 1**), the rate of ATP production in the glycolytic pathway (glycoATP Production Rate) is equivalent to Glycolytic Proton Efflux Rate (glycoPER, **Equation 2**) and can be calculated using the same approach previously validated for the Agilent Seahorse XF Glycolytic Rate Assay (for more details see **White paper: Improving Quantification of Cellular Glycolytic Rate Using Agilent Seahorse XF Technology**)

$$\text{glycoATP Production Rate (pmol ATP/min)} = \text{glycoPER (pmol H}^+/\text{min)} \quad (\text{Equation 2})$$

Mitochondrial ATP production rate calculation

The rate of oxygen consumption that is coupled to ATP production during OXPHOS can be calculated as the OCR that is inhibited by addition of the ATP synthase inhibitor, oligomycin (**Equation 3**):

$$\text{OCR}_{\text{ATP}} \text{ (pmol O}_2\text{/min)} = \text{OCR (pmol O}_2\text{/min)} - \text{OCR}_{\text{Oligo}} \text{ (pmol O}_2\text{/min)} \quad (\text{Equation 3})$$

Transformation of **OCR_{ATP}** to the rate of mitochondrial ATP production consists of: multiplying by 2 to convert molecules of O₂ to oxygen (O) atoms consumed, then multiplying the P/O ratio, the number of molecules of ADP phosphorylated to ATP per atom of O reduced by an electron pair flowing through the electron transfer chain (**Equation 4** on page 8).

1 Introduction

Mitochondrial ATP production rate calculation

The Seahorse XFp Real-Time ATP Rate Assay uses an average P/O value of 2.75 for these calculations that was validated to accurately represent the assay conditions for a broad panel of cells under different fuels availabilities (see [White Paper: Quantifying Cellular ATP Production Rate Using Agilent Seahorse XF Technology](#)).

With these considerations, the rate of mitochondrial ATP production is calculated according to [Equation 4](#):

$$\text{mitoATP Production Rate (pmol ATP/min)} = \text{OCR}_{\text{ATP}} \text{ (pmol O}_2\text{/min)} * 2 \text{ (pmol O/pmol O}_2\text{)} * \text{P/O (pmol ATP/pmol O)} \quad \text{(Equation 4)}$$

Finally, the total cellular ATP Production Rate ([Equation 5](#)) is the sum of the glycolytic and mitochondrial ATP production rates:

$$\text{ATP Production Rate (pmol ATP/min)} = \text{glycoATP Production Rate (pmol ATP/min)} + \text{mitoATP Production Rate (pmol ATP/min)} \quad \text{(Equation 5)}$$

Glossary

- **glycoATP Production Rate:** Rate of ATP production (expressed in pmol ATP/min) associated with the conversion of glucose to lactate in the glycolytic pathway.
- **mitoATP Production Rate:** Rate of ATP production (expressed in pmol ATP/min) associated with oxidative phosphorylation in the mitochondria.
- **Total ATP Production Rate:** The sum of mitochondrial ATP and glycolytic ATP production rates in live cells under appropriate assay conditions.
- **P/O ratio:** Number of molecules of ATP synthesized (that is, moles of ADP phosphorylated to ATP) per atom of oxygen (O) reduced by an electron pair flowing through the mitochondrial electron transport chain.
- **XF ATP Rate Index:** Ratio of the mitoATP Production Rate divided by glycoATP Production Rate at a certain time point. This is a valuable metric for detecting changes and/or differences in metabolic phenotype. An increase in the XF ATP Rate Index represents a more oxidative / less glycolytic phenotype and *vice-versa*.
- **Induced Assay:** An XFp Real-Time ATP Rate Assay workflow that includes an acute injection of an experimental compound prior to serial injections of oligomycin and rotenone/antimycin A. This workflow allows for measurement of *in situ* activation or repression of ATP production, as well as real-time shifts in the XF ATP Rate Index (mitoATP Production Rate / glycoATP Production Rate).
- **Buffer Factor:** Buffer capacity of the measurement system, comprising the assay medium and XF assay conditions (instrument, sensor, labware)
- **Glycolysis:** In the context of Seahorse XF Assays, the process of converting glucose to lactate.
- **Proton Efflux Rate:** The number of protons exported by cells into the assay medium over time, expressed as pmol/min.
- **Glycolytic Proton Efflux Rate:** Proton Efflux rate derived from glycolysis (discounting the effect of CO₂-dependent acidification). This measurement is highly correlated with the extracellular lactate production rate.
- **ATP coupled respiration:** The portion of basal respiration that is being used to drive ATP production. It is quantified by injection of the ATP synthase inhibitor oligomycin.

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Kit Contents

The Seahorse XFp Real-Time ATP Rate Assay Kit includes six foil pouches, each containing reagents sufficient for one complete XFp Real-Time ATP Rate Assay in an 8-well Agilent Seahorse XFp Cell Culture Miniplate.

Every pouch includes one tube of Oligomycin and one tube of a mixture of Rotenone + Antimycin A (see **Table 1**).

Table 1 Seahorse XFp Real-Time ATP Rate Assay Kit Contents

Comound	Target	Cap color	Quantity per tube
Oligomycin	ATP synthase inhibitor (complex V)	Blue	12.6 nmol
Rotenone + Antimycin A (Rot/AA)	Mitochondrial ETC complexes I and III, respectively	Red	5.4 nmol each

Kit Shipping and Storage

Product ships at ambient temperature and can be stored at room temperature. Products are stable for 18 months from the date of manufacture (listed on the box).

2 Kit Information

Additional Agilent Products Required

Additional Agilent Products Required

The following Agilent products are also required for performing the Seahorse XFp Real-Time ATP Rate Assay, but not supplied with the kit. For a complete list of materials required to perform an XF assay, please visit Basic Procedures to Run an XF Assay on an XFp Analyzer at

[http://www.agilent.com/en-us/products/cell-analysis-\(seahorse\)/basic-procedures-to-run-an-xf-assay](http://www.agilent.com/en-us/products/cell-analysis-(seahorse)/basic-procedures-to-run-an-xf-assay)

Item	Vendor	Part number
Seahorse XFp Analyzer	Agilent Technologies	
Seahorse XF DMEM Medium, pH 7.4 or Seahorse XF RPMI Medium, pH 7.4	Agilent Technologies	103575-100
	Agilent Technologies	103576-100
Seahorse XFp FluxPak	Agilent Technologies	103022-100
Seahorse XF 1.0 M Glucose solution	Agilent Technologies	103577-100
Seahorse XF 100 mM Sodium Pyruvate solution	Agilent Technologies	103578-100
Seahorse XF 200 mM L-Glutamine solution	Agilent Technologies	103579-100

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Assay Workflow

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Day of Assay 15

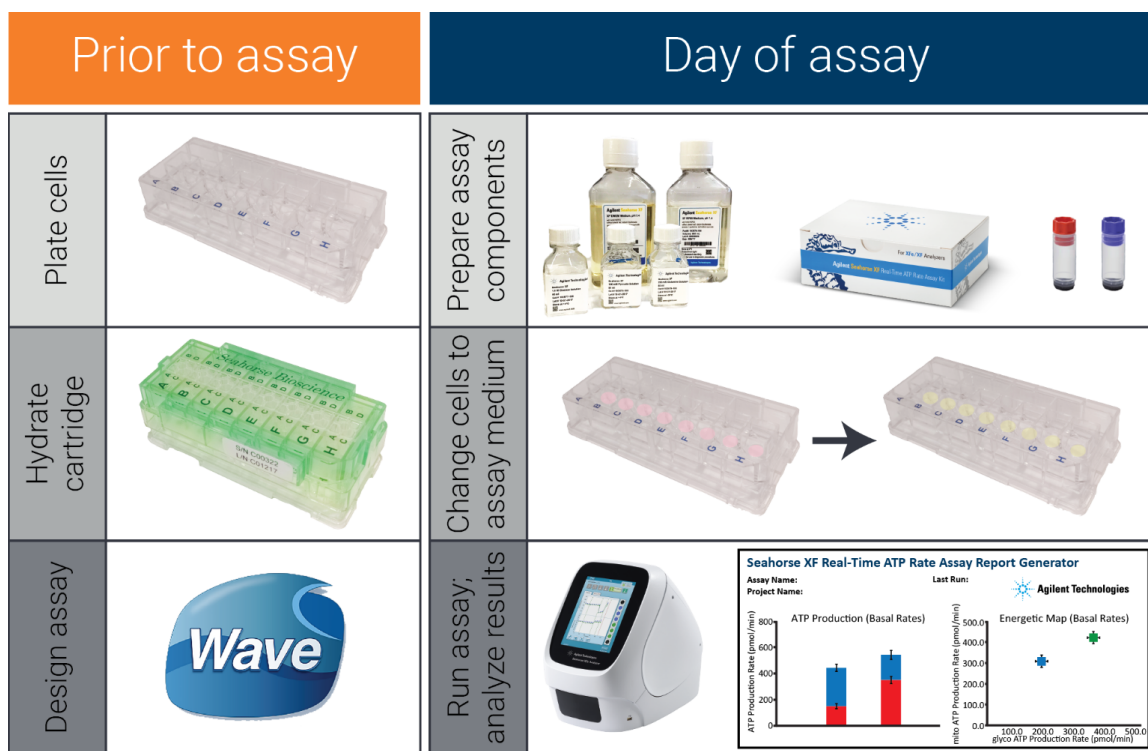


Figure 2. Agilent Seahorse XFe Real-Time ATP Rate Assay workflow

Day Prior to Assay

- 1 Turn on the Agilent Seahorse XFp Analyzer, and let it warm up overnight (minimum five hours).
- 2 For adherent cells, plate cells at a predetermined density in the Seahorse XFp Microplate using the appropriate cell culture growth medium. Refer to Basic Procedures to Run an XF Assay on an XFp Analyzer at [http://www.agilent.com/en-us/products/cell-analysis-\(seahorse\)/basic-procedures-to-run-an-xf-assay](http://www.agilent.com/en-us/products/cell-analysis-(seahorse)/basic-procedures-to-run-an-xf-assay) for details.

For suspension cells, see section: **“Prepare Seahorse XFp cell culture miniplate for assay”** on page 15.
- 3 Hydrate a sensor cartridge at 37 °C in a non-CO₂ incubator overnight. (Refer to Basic Procedures).
- 4 Design experiment in Wave or use ATP Production Rate Assay Template (Standard or Induced Assay). Make any necessary group modifications to the template for the specific assay design.

Day of Assay

Prepare cartridge

- 1 Remove the sensor cartridge from the incubator and discard water from the utility plate. Replace with prewarmed XF Calibrant.
- 2 Place the sensor cartridge onto the utility plate and incubate for 45-60 minutes in a non-CO₂ 37 °C incubator prior to load the ports.

Prepare assay medium

- 1 Under sterile conditions, prepare assay media by supplementing 10 mL of Seahorse XF DMEM Medium, pH 7.4 with 10 mM of XF glucose, 1 mM of XF pyruvate, 2 mM of XF glutamine. These are recommended initial conditions; however, the desired assay medium composition can be varied depending on cell type or *in vitro* culture conditions.
- 2 Warm assay medium to 37 °C.
- 3 Incubate at 37 °C until ready to use.

NOTE

If the assay medium is significantly altered from this formulation (that is, higher concentrations of XF glucose, XF pyruvate or XF glutamine and/or other media additives), the pH of the assay medium should be checked and adjusted to 7.4 if needed, and the Buffer Factor Protocol must be performed to derive the buffer factor value. Consult Seahorse XF Buffer Factor Protocol User Guide.

Prepare Seahorse XFp cell culture miniplate for assay

For adherent cells,

- 1 Remove cell culture miniplate from 37 °C CO₂ incubator and examine cells under microscope to confirm consistent plating and proper cell morphology.
- 2 Wash cells (refer to Basic Procedures for more details). Remove the cell culture growth medium in the cell culture miniplate. Wash once with warmed assay medium using a multichannel pipette and incubate with assay medium at 37 °C in a non-CO₂ incubator for 45-60 minutes prior to the assay.

3 Assay Workflow

Prepare stock compounds

- 3 Before starting the XF assay, remove the assay medium AGAIN, and add fresh, warm assay medium to each well (see **Table 4** on page 18 for appropriate starting well volumes).

For suspension cells,

- 1 Pellet cells out of their growth medium, and resuspend in warm assay medium.
- 2 Count cells and suspend at a concentration such that seeding 50 μL of cells contains the desired cell number per well, leaving four wells without cells as background correction wells.
- 3 Add desired cells/well, then centrifuge gently to adhere.
- 4 Gently add assay medium to each well. Total well volume should match the appropriate Starting Well Volume as indicated in **Table 4** on page 18.
- 5 Incubate the plate at 37 $^{\circ}\text{C}$ in a non- CO_2 incubator for 45-60 minutes prior to the assay.

Prepare stock compounds

NOTE

Use compounds the same day they are reconstituted. Do not freeze. Discard any remaining compound.

- 1 Remove one foil pack from the kit box, then open pouch and remove the Oligomycin (blue cap) and Rotenone/Antimycin A (red cap) vial.
- 2 Tap down the vials to ensure powder is on the bottom of the tube before opening the vials.
- 3 Resuspend each component with the appropriate volume of prepared assay medium as described in **Table 2**. Vortex ~one minute to ensure full resuspension of compounds.

Table 2 Stock solutions

Compound	Volume of assay medium	Resulting stock concentration
Oligomycin	168 μL	75 μM
Rotenone + Antimycin A	216 μL	25 μM

3 Assay Workflow

Prepare compounds for loading in sensor cartridge

Prepare compounds for loading in sensor cartridge

Prepare 0.3 mL of each compound in assay medium as described in **Table 3**. It is recommended to use 1.5 μM of oligomycin and 0.5 μM Rotenone + Antimycin A (final concentration).

Table 3 Compound preparation for performing the XFp Real-Time ATP Rate Assay on an XFp Analyzer

	Stock volume (μL)	Medium volume (μL)	10X [Port] (μM)	[Final well] (μM)
Port A Oligomycin	60	240	15	1.5
Port B Rot/AA	60	240	5	0.5

Load sensor cartridge

- **Standard Assay** - No acute injection before oligomycin and rotenone/antimycin A. Load compounds into the following ports of a hydrated sensor cartridge:
Port A: Oligomycin
Port B: Rotenone/antimycin A
- **Induced Assay** -To inject a test compound(s), use port(s) A for the desired compound(s) and then load ports as follows:
Port A: experimental compound (acute injection) or vehicle control
Port B: Oligomycin
Port C: Rotenone/antimycin A

Table 4 on page 18 lists the appropriate volumes and concentrations for injection schemes using two or more ports.

3 Assay Workflow

Load template onto the Seahorse XFp Analyzer

Table 4 Starting well assay medium and compound injection volumes for Agilent Seahorse XFp Analyzers.

Starting well volume: 180 µL assay medium		
Port	Add to port volume	Port concentration
A	20 µL	10X
B	22 µL	10X
C	25 µL	10X
D	27 µL	10X

Load template onto the Seahorse XFp Analyzer

(If template is already present, skip this step.)

Personal computer (internet access required)

- 1 Download the Seahorse XFp Real-Time ATP Rate Assay Report Generator from the Agilent Website. Both XFp Real-Time ATP Rate Assay Standard and Induced Assay Templates are included in the download folder.

NOTE

Select the appropriate Seahorse XFp Analyzer (Seahorse XFp) when registering to download the Report Generator and accompanying Assay Templates.

- 2 Transfer to a USB drive.

Seahorse XFp Analyzer

- 1 Insert USB drive in front USB port of the Seahorse XFp Analyzer
- 2 Click **Settings**, then click **Template Management**.
- 3 Click the **USB** tab, and locate the Assay template(s) to upload.
- 4 Click the checkbox next to each Assay Template to be imported, then click **Import**.
- 5 All imported Assay template(s) will now be available for selection from the 'Local' tab when starting an XFp assay.

3 Assay Workflow

Running the XFp Real-Time ATP Rate Assay

Running the XFp Real-Time ATP Rate Assay

- 1 Click **Start**.
- 2 Select the **Seahorse XFp Real-Time ATP Rate Assay** or **Seahorse XFp Real-Time ATP Rate Assay (Induced Assay)** template from the Local tab.
- 3 **Group Page**: confirm or modify the default groups and plate map, then click the right arrow.
- 4 **Protocol Page**: confirm or modify the Instrument Protocol for additional measurements cycles during the assay then click the right arrow.
- 5 **Summary Page**: Change the name of the assay results file, if desired. Click **Start Run** when ready.
- 6 When prompted, place the loaded sensor cartridge with the utility plate into the Seahorse XFp Analyzer, then click **Continue**. Calibration takes approximately 20 minutes.

NOTE

Remove the cartridge lid and verify correct plate orientation.

- 7 After completing the Calibration, remove the utility plate and place the Seahorse XFp Cell Culture Miniplate on the tray and click **Continue** to start the assay.

3 Assay Workflow

Using the Agilent Seahorse XF Real-Time ATP Rate Assay Report Generator

Using the Agilent Seahorse XF Real-Time ATP Rate Assay Report Generator

The Agilent Seahorse XF Real-Time ATP Rate Assay Report Generator automatically calculates the XF Real-Time ATP Rate Assay Parameters (mitoATP Production Rate, glycoATP Production Rate, total ATP Production Rate, XF ATP Rate Index, % glycolysis, and % OXPHOS). The Seahorse XF Real-Time ATP Rate Assay Report Generator can be used with either a standard or an induced assay protocol, and provides a convenient one-page assay summary.

The Report Generator can be download from Agilent website. Visit https://www.agilent.com/cs/library/usermanuals/public/Report_Generator_User_Guide_Seahorse_XF_Real_Time_ATP_Rate_Assay.pdf to learn more about the Seahorse XF Real-Time ATP Rate Assay Report Generator and download the User Guide.

- 1 After finishing the assay, insert an USB Drive on the from USB of the Seahorse XFp Analyzer and save your assay file (*.asyr)
- 2 Open your file in Wave Desktop (2.4 or higher).
- 3 Open Overview Tab and look at OCR and PER Data. Confirm that Assay Media and Buffer Factor is assigned to every group including background wells.
- 4 Click on **Export tab** and select **Microsoft Excel** to export the completed run as an .xlsx file.
- 5 Load the data file in the XF Real-Time ATP Rate Assay Report Generator and select groups to display. If the assay has more than three injections, select if there is an acute injection and which injection corresponds to Oligomycin. Rot/AA injection will be automatically assigned as the immediate following injection to oligomycin.
- 6 Click **Update Summary** to obtain XF Real-Time ATP Rate Assay Report. For further details, consult the [Agilent Seahorse XF Real-Time ATP Rate Assay Report Generator User Guide](#).

Frequently Asked Questions

How is the mitoATP Production Rate calculated if there are different oxidizable substrates (with different P/O ratios) in the assay media?

To convert ATP-coupled respiration (OCR) into mitochondrial ATP production rates, the stoichiometry of ADP phosphorylated to ATP per atom of oxygen reduced at the terminal point of electronic transport chain must be determined. The maximum theoretical P/O value varies between different fuels and depends on the stoichiometry/pathway of the fuel oxidation, as well as the efficiency of the F_1F_0 ATP synthase. Under standard XF experimental conditions, cells are provided with a mixture of fuels (predominantly glucose, pyruvate and glutamine), and often endogenous fuel stores (glycogen, fatty acids, other amino acids) are available for mitochondrial oxidation. Thus, it was tested with several cell lines and validated that a P/O value of 2.75 accurately represents the average P/O ratio of the mixture of exogenous and endogenous oxidizable substrates. For more information, see the white paper “Quantifying Cellular ATP Production Rate Using Agilent Seahorse XF Technology” at:

<https://seahorseinfo.agilent.com/acton/fs/blocks/showLandingPage/a/10967/p/p-0157/t/page/fm/1>

When an Induced XF Real-Time ATP Rate assay is performed, how is the Induced rate calculated and reported?

During an XF Real Time ATP Rate Induced Assay, the Induced Rate(s) displayed in the Summary Printout and Average Assay Parameter Calculations of the Measure Sheet are calculated *as the average rate of all the measurements between the acute injection and the oligomycin injection*. However, induced rates at each time point can be obtained from the Kinetic Rate Data (Measure Sheet).

What is the XF ATP Rate Index as defined the in XFp Real-Time ATP Rate Assay?

The XF ATP Rate Index is the ratio of the mitoATP Production Rate to the glycoATP Production Rate (i.e., mitoATP rate / glycoATP rate) This ratio represents a quantitative metric of the cellular metabolic phenotype. Metabolic indices greater than 1.0 represent cellular metabolism where greater than 50 % of cellular ATP is derived from mitochondrial through ETC/OXPHOS, while indices less than 1 indicate greater than 50 % of total ATP is produced by the glycolytic pathway. Since the metabolic index is a ratio-metric measurement, it is highly sensitive to changes or shifts in metabolic phenotype.

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Why is the XF ATP Rate Index different for the same cell type when the cells are plated at different densities?

The metabolic phenotype of the cell will be influenced by the cellular demand for ATP. Cells that are proliferating or differentiating have, in general, higher glycolytic rates than cells that are confluent (slow growing) or terminally differentiated. In order to compare results between experiments it is recommended to maintain consistent cell culture conditions and cell seeding densities throughout the investigation.

Why must Seahorse XF DMEM Medium, pH 7.4 or Seahorse XF RPMI Medium, pH 7.4 be used to perform the assay?

The calculation of the glycoATP Production Rate requires an absolute measurement of the glycolytic proton efflux rate during the XF Real-Time ATP Rate Assay. To properly calculate PER, the assay media must have a fixed buffer capacity. A low concentration of HEPES in the media provides consistent buffer capacity values across the time frame of the assay. Although addition of HEPES buffer reduces the ECAR signal slightly, it significantly improves the quality and consistency of the ECAR data, as well as the accuracy of the transformation to PER (For more details see [White paper: Improving Quantification of Cellular Glycolytic Rate Using Agilent Seahorse XF Technology](#)). Additionally, the use of XF DMEM or XF RPMI medium with the pH pre-adjusted to 7.4 saves time in assay preparation and ensures a consistent assay media pH value across XF experiments.

Can I use other Seahorse XF assay media to run the XF Real-Time ATP Rate Assay?

Calculation of accurate glycoATP production rates requires the use of an assay medium without phenol red or bicarbonate and a low concentration of HEPES buffer. Thus, the use of Agilent Seahorse XF DMEM (or RPMI), pH 7.4 is strongly recommended. Any deviation from the recommended medium and supplements (glucose, pyruvate, glutamine) will require that the Buffer Factor should be determined empirically (using the XF Analyzer) for each assay (See the Seahorse XF Buffer Factor Protocol for further information). Any assay medium containing phenol red cannot be used for the XF Real-Time ATP Rate Assay.

When running an induced XF Real-Time ATP Rate Assay, the OCR after oligomycin injection is higher than the basal OCR, what happened?

Addition of compounds that uncouple electron transport from oxidative phosphorylation (like FCCP, DNP, etc.) before the oligomycin injection will typically result in an increase in OCR. However, this respiration is not coupled to ATP production since the mitochondrial membrane potential is decreased or dissipated without participation of the ATP synthase, so little to no ATP is

4 Frequently Asked Questions

generated under these circumstances. In these cases, the basal mitoATP Production Rate cannot be calculated accurately. If an uncoupler compound is suspected, then it is recommended to include a control group with an injection of assay medium + vehicle to calculate basal mitoATP Production Rates.

4 Frequently Asked Questions

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